



King's Research Portal

DOI:

[10.3389/fimmu.2020.622442](https://doi.org/10.3389/fimmu.2020.622442)

Document Version

Peer reviewed version

[Link to publication record in King's Research Portal](#)

Citation for published version (APA):

Willsmore, Z. N., Harris, R. J., Crescioli, S., Hussein, K., Kakkassery, H., Thapa, D., Cheung, A., Chauhan, J., Bax, H. J., Chenoweth, A., Laddach, R., Osborn, G., McCraw, A., Hoffmann, R. M., Nakamura, M., Geh, J. L., MacKenzie-Ross, A., Healy, C., Tsoka, S., ... Karagiannis, S. N. (2021). B Cells in Patients With Melanoma: Implications for Treatment With Checkpoint Inhibitor Antibodies. *Frontiers in Immunology*, 11, [622442]. <https://doi.org/10.3389/fimmu.2020.622442>

Citing this paper

Please note that where the full-text provided on King's Research Portal is the Author Accepted Manuscript or Post-Print version this may differ from the final Published version. If citing, it is advised that you check and use the publisher's definitive version for pagination, volume/issue, and date of publication details. And where the final published version is provided on the Research Portal, if citing you are again advised to check the publisher's website for any subsequent corrections.

General rights

Copyright and moral rights for the publications made accessible in the Research Portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognize and abide by the legal requirements associated with these rights.

- Users may download and print one copy of any publication from the Research Portal for the purpose of private study or research.
- You may not further distribute the material or use it for any profit-making activity or commercial gain
- You may freely distribute the URL identifying the publication in the Research Portal

Take down policy

If you believe that this document breaches copyright please contact librarypure@kcl.ac.uk providing details, and we will remove access to the work immediately and investigate your claim.

B cells in patients with melanoma: implications for treatment with checkpoint inhibitor antibodies

Zena N Willsmore¹, Robert J Harris¹, Silvia Crescioli¹, Khuluud Hussein¹, Helen Kakkassery¹, Deepika Thapa¹, Anthony Cheung^{1,2}, Jitesh Chauhan¹, Heather J Bax^{1,3}, Alicia Chenoweth^{1,2}, Roman Laddach^{1,4}, Gabriel Osborn¹, Alexa McCraw¹, Ricarda M Hoffmann¹, Mano Nakamura¹, Jenny L C Geh⁵, Alastair MacKenzie-Ross⁵, Ciaran Healy⁵, Sophia Tsoka⁴, James F Spicer³, Sophie Papa^{6,7}, Linda Barber⁸, Katie E Lacy¹, Sophia N Karagiannis^{1,2}

¹ St. John's Institute of Dermatology, School of Basic & Medical Biosciences, King's College London, Tower Wing, 9th Floor, Guy's Hospital, London, SE1 9RT, United Kingdom

² Breast Cancer Now Research Unit, School of Cancer & Pharmaceutical Sciences, King's College London, Guy's Cancer Centre, London, United Kingdom

³ School of Cancer & Pharmaceutical Sciences, King's College London, 3rd Floor, Guy's Hospital, London, United Kingdom

⁴ Department of Informatics, Faculty of Natural and Mathematical Sciences, King's College London, London, UK.

⁵ Department of Plastic Surgery at Guy's, King's, and St. Thomas' Hospitals, London, United Kingdom

⁶ Department of Medical Oncology, Guy's and St Thomas' NHS Foundation Trust, London, United Kingdom

⁷ ImmunoEngineering, School of Cancer and Pharmaceutical Sciences, Faculty of Life Sciences and Medicine, King's College London, London, UK

⁸ School of Cancer & Pharmaceutical Sciences, King's College London, Room 1.21 Henriette Raphael House, Guy's Campus, London, United Kingdom

Running title:

B cell responses in melanoma immunotherapy

Correspondence to:

Sophia Karagiannis, St. John's Institute of Dermatology, School of Basic & Medical Biosciences, King's College London, Tower Wing, 9th Floor, Guy's Hospital, London, SE1 9RT, United Kingdom

Email: sophia.karagiannis@kcl.ac.uk

Funding:

The authors acknowledge support by the Cancer Research UK King's Health Partners Centre at King's College London (C604/A25135); The Guy's and St Thomas's Foundation Trust Charity Melanoma Special Fund; CRUK/NIHR in

England/DoH for Scotland, Wales and Northern Ireland Experimental Cancer Medicine Centre (C10355/A15587); Breast Cancer Now (147; KCL-BCN-Q3); the Medical Research Council (MR/L023091/1); Cancer Research UK (C30122/A11527; C30122/A15774). The research was supported by the National Institute for Health Research (NIHR) Biomedical Research Centre (BRC) based at Guy's and St Thomas' NHS Foundation Trust and King's College London (IS-BRC-1215-20006). The authors are solely responsible for study design, data collection, analysis, decision to publish, and preparation of the manuscript. The views expressed are those of the author(s) and not necessarily those of the NHS, the NIHR, or the Department of Health.

Conflicts of Interest:

SNK and JFS are founders and shareholders of Epsilogen Ltd. HJB is now employed through a fund provided by Epsilogen Ltd. All other authors declare no conflicts of interest.

Abstract

The contributions of the humoral immune response to melanoma are now widely recognised, with reports of positive prognostic value ascribed to tumour-infiltrating B cells (TIL-B) and increasing evidence of B cells as key predictors of patient response to treatment. There are disparate views as to the pro- and anti-tumour roles of B cells. B cells appear to play an integral role in forming tumour-associated tertiary lymphoid structures (TLS) which can further modulate T cell activation. Expressed antibodies may distinctly influence tumour regulation in the tumour microenvironment, with some isotypes associated with strong anti-tumour immune response and others with progressive disease. Recently, B cells have been evaluated in the context of cancer immunotherapy. Checkpoint inhibitors (CPI), targeting T cell effector functions, have revolutionised the management of melanoma for many patients, however, there remains a need to accurately predict treatment responders. Increasing evidence suggests that B cells may not be simple bystanders to CPI immunotherapy. Mature and differentiated B cell phenotypes are key positive correlates of CPI response. Recent evidence also points to an enrichment in activatory B cell phenotypes and the contribution of B cells to TLS formation may facilitate induction of T cell phenotypes required for response to CPI. Contrastingly, specific B cell subsets often correlate with immune-related adverse events (irAEs) in CPI. With increased appreciation of the multifaceted role of B cell immunity, novel therapeutic strategies and biomarkers can be explored and translated into the clinic to optimise CPI immunotherapy in melanoma.

Introduction

During early stages, primary melanoma lesions are removable through surgical intervention that is largely curative. In advanced disease however, melanoma can spread to regional lymph nodes and metastasise to distant sites. Historically treatment options were limited in advanced disease to palliative cytotoxic chemotherapy, leading to poor 5-year survival rates. Over the last decade immunotherapy, driven by checkpoint inhibitor antibodies, has transformed patient prognosis.

The rationale behind the use of checkpoint inhibitor therapy lies in the highly immunogenic nature of melanoma (1,2). Melanoma carries a large mutational load, providing a range of tumour-specific antigens that are thought to elicit a host immune response. However, melanoma cells can evade immunosurveillance via activation of different immune-inhibitory pathways, including via immune checkpoint molecules and their downstream signals. Physiologically, checkpoint pathways play a role in immune homeostasis, providing negative feedback stimuli to prevent autoimmune reactivity. The best-studied checkpoint pathways are those of negative regulatory molecules cytotoxic T lymphocyte associated protein-4 (CTLA-4) and of the Programmed Death Receptor 1 (PD-1) and its ligands PD-L1 and PD-L2.

Checkpoint inhibitors (CPI) were designed to promote immune-mediated elimination of tumour cells through modulation of T cell responses. Anti-CTLA-4 (Ipilimumab approved in 2011) (3) or anti-PD-1 (Pembrolizumab and Nivolumab approved in 2014) (3,4) antibodies inhibit binding of checkpoint molecules with their respective ligand in the tumour microenvironment or draining lymph nodes. A newer anti-PD-L1 agent, Atezolizumab, was approved by the FDA in July 2020 (5). CPI are approved for use as monotherapy in metastatic melanoma and as adjuvant therapy. More recently combination Nivolumab and Ipilimumab have been approved for metastatic disease in select patient groups. Their clinical efficacy and long-term outcomes are highlighted in the CheckMate 067 clinical trial (ClinicalTrials.gov NCT01844505) (4). In 945 patients with stage III or IV melanoma, this trial directly evaluated combination therapy with Nivolumab plus Ipilimumab in comparison with monotherapy with Nivolumab or with Ipilimumab. At a minimum follow-up of 60 months, the median overall survival (OS) was more than 60.0 months (median not reached) in the Nivolumab-plus-Ipilimumab group, 36.9 months in the Nivolumab group and 19.9

months in the Ipilimumab group. The 5-year OS rates were 52%, 44%, and 26% respectively (4).

Despite the undeniable clinical success of CPI therapy several challenges remain. There is currently a lack of biomarkers, limiting our ability to predict who will respond to treatment. Another important consideration is recognising who will develop toxicities, that are unfortunately common particularly with anti-CTLA-4 treatment and in the context of combination anti-CTLA-4 and anti-PD-1 therapy as compared with monotherapy. These toxicities, known as immune related adverse events (irAE) vary in severity, can affect any organ system but most commonly target the skin, the gastrointestinal and the endocrine systems (6). Although many of these toxicities are manageable/reversible they frequently lead to treatment disruption or discontinuation. Therefore, understanding the mechanisms underlying irAE and predicting these irAE is a much-needed clinical strategy.

The role of T cells in CPI therapy has been extensively reviewed in literature. Contrastingly, B cell immunity has been less studied. B cells have a wide range of roles, including critical functions as professional antigen presenting cells (APCs) and they are capable of secreting cytokines and antibodies which enable them to conduct antibody-dependent cell-mediated cytotoxicity (ADCC), antibody-dependent cellular phagocytosis (ADCP) and complement-dependent cytotoxicity (CDC). Considering their comprehensive functions and with B cells making up one of the two arms of the adaptive immune system, it is not surprising to envisage a role within melanoma immunity and response to CPI. However, the exact role B cells play in tumour immunity is unclear with the existence of different cell subsets with juxtaposing functions, such as activatory and regulatory B cells (Breg), and the presence of B cells in tertiary lymphoid structures (TLS). The potential pro- and anti-tumour roles of B cells in melanoma are summarised in Figure 1. In this review we discuss emerging evidence for the significance of B cell and antibody immune responses in melanoma and specifically in the context of CPI therapy.

The CTLA-4 and PD-1 pathways

CTLA-4 is an inhibitor of T cell activation primarily expressed on naïve T cells after activation (7–10) and constitutively expressed on FoxP3+ regulatory T cells (Treg)

(10). T cell activation is dependent not only T cell receptor (TCR) binding with an antigen presented via an APC, but also on the presence of a costimulatory second signal, typically through binding of CD28 on the T cell to CD80/86 on the APC. CTLA-4 is a competitive CD28 homolog that has a higher affinity to CD80 (B7-1), and to a lesser degree CD86 (B7-2) than CD28 therefore inhibiting T cell co-stimulation (10). TCR signalling up-regulates cell surface CTLA-4 expression, reaching peak expression at 2 to 3 days after activation (7,8). This provides a negative feedback loop for T cell activation.

PD-1 is also a member of the immunoglobulin superfamily known to contribute to immune homeostatic processes by delivering inhibitory signals upon engagement with its ligands, programmed death ligands 1 and 2 (PD-L1/2). Like CTLA-4, PD-1 is thought to be a negative regulator of T cell function, regulating peripheral tolerance and T cell responses. PD-1 is expressed more broadly than CTLA-4 and can be found on T cells, B cells, natural killer (NK) cells and a variety of peripheral tissues (11). Expression of PD-1 is upregulated upon activation of T cells and B cells (12). The ligand PD-L1 is broadly expressed by immune cells including T cells, B cells, dendritic cells and macrophages, and in non-lymphoid tissues including on tumour cells or stromal elements in the tumour microenvironment (TME) (11,13). Recent evidence suggests that CD28 is also a primary target for PD-1 (14,15).

Intracellularly, both PD-1 and CTLA-4 signal via SHP2 and converge to inhibit downstream P13K signalling (16,17). Although it is reported that B cells express PD-1 and PD-L1, these cells have not been recognised as a primary target for anti-PD-1 therapy. CTLA-4 also acts extracellularly to clear the CD28 ligands CD80/86 from surrounding cells, including APCs, by trans-endocytosis *in vivo*, further inhibiting T-cell activation (18). While B cells are also known to express CTLA-4 ligands CD80/CD86, again they are not considered the key target cells of anti-CTLA-4 therapy (10).

Overview of B cells in melanoma, in the tumour microenvironment and tertiary lymphoid structures

Besides constituting an amalgamation of malignant cells, the tumour microenvironment (TME) can be thought of as a distinct complex “organ” which contains an array of different cells, such as immune cells, tumour-associated vasculature and lymphatics, fibroblasts and pericytes (15). The immune cell infiltrates

are of special importance as they often correlate with anti-tumour activity and have been reported to be associated with a more favourable prognosis in some settings. Melanoma is a classically highly immunogenic tumour (19) and lymphocyte infiltrates in general have been shown to have positive prognostic value in patients with primary cutaneous melanoma (20). Immune cell subsets which are correlated with increased patient survival outcome include CD8⁺ memory T cells, CD4⁺ T-helper cells (21), B cells (22) and NK cells (23). Immune cell infiltrates harbour the potential to exert cytotoxic anti-tumour activity, which may be exacerbated by checkpoint inhibitor immunotherapy.

Lymphocyte activation typically occurs in secondary lymphoid structures which include lymph nodes, the spleen and mucosal-associated lymphoid tissue (24). During chronic inflammation, TLS may transiently form, which confer sites enriched in B and T lymphocytes and dendritic cells present from disordered compositions to more structured and often highly specialised germinal centre-like formations (25). As tumours share many of the characteristics of chronic inflammation, it is unsurprising that TLS have been identified in a range of cancers adjacent to the tumour site, including in 26% of metastatic melanomas (26). Importantly, the presence of TLS has been shown to be associated with a more favourable prognosis (27,28) and the promotion of immunotherapy response (discussed below) (29). There is compelling evidence of ongoing B cell maturation in these sites, driving clonal amplification, class switching and somatic hypermutation (SMH). It is therefore clear that intralesional B lymphocyte structural organisation and the interaction with other lymphocytic cell subsets within TLS could be of particular importance to overall B cell-mediated anti-tumour activity.

The majority of melanoma lesions contain significant populations of B cells, usually localised within the tumour stroma (one study has reported that 86% of primary melanomas harbour at least ten CD20⁺ stromal B cells per mm² tumour (30). Furthermore, tumour infiltrating B lymphocytes TIL-B in primary lesions make up an average of 10% of the total infiltrating lymphocytes (31–33) and can be found adjacent to various other immune cell subsets. CD22⁺ B cells are enriched in melanoma lesions compared to healthy skin, and CD20 and CD22 mRNA expression is further enhanced with metastasis (34,35), indicating the presence of a pronounced humoral immune response in melanoma.

These TIL-B have been shown to contribute to anti-tumour immunity via antibody responses to melanoma-associated antigens. In one study, 28% of melanoma patient-derived B cell cultures were capable of binding to a wide range of antigens expressed by melanoma cell lines, as opposed to just 2% of those derived from healthy individuals, and this was shown to further increase with metastatic disease (36). In concordance, plasma cells in melanoma lesions were shown to be polyclonal and to predominantly express IgG and IgA (37) implicating antigen reactivity and class switching, and thus a possible melanoma-reactive B cell immune response. Secreted immunoglobulins, especially of the IgG1 subclass, may be able to exert potent tumoricidal effects via ADCC, as demonstrated *in vitro* using a patient-derived monoclonal antibody (36). ADCC however is just one of several effector mechanisms mediated by secreted immunoglobulin. A neutralising anti-human growth factor (HGF) antibody has been shown to suppress tumour growth *in vitro* (38). B cells may also act as antigen presenting cells, contributing to T-helper cell activation and anti-tumour effector mechanisms (39). Finally, activatory B cells may facilitate anti-tumour immunity via the release of pro-inflammatory cytokines such as IL-2, TNF α and IFN γ (40,41). TNF α produced by B cells may control the development of follicular dendritic cells, the formation of B cell follicles and T cell-dependent antibody responses (42). Collectively, several studies therefore support the presence of an antigen-specific B cell immune response to melanoma.

In accordance with expectations arising from these anti-tumour properties, the percentage of tumour-infiltrating and peritumoral B cells have been found to positively correlate with more favourable patient survival in the vast majority of melanoma cohort studies (22,43,44). However, some studies have in contrast found no significant association between B cell infiltration and overall patient survival (31), and even a correlation of B cell infiltrates with a poorer prognosis (33). These discrepancies may arise from differences in experimental design and analyses (e.g. using raw B cell counts *versus* percentage of infiltrating populations) or may reflect the confounding presence of immunosuppressive regulatory B cells.

B cell subsets in melanoma and the role of Bregs

Several studies have demonstrated the enriched presence of various differentiated B cell phenotypes such as memory, plasmablasts (PB) and plasma cells within melanoma tumours. Elevated percentages of PBs (short-lived cells and part of the

rapid antigen response) have been reported in the circulation of patients with melanoma compared to blood from healthy individuals. PB are short-lived cells and part of a rapid antigen response. Although PB have a potentially positive role in melanoma, the functional plasticity of these cells may result in transient switching to a “regulatory” pro-tumour phenotype owing to expression of TGF β and IL-10 by this cell subset (41).

IL-10-producing “regulatory” B (B10) cells (45) have been shown to be functional in murine models of tolerance (46), in preventing chronic inflammation (47–50) and in curbing anti-tumour immune responses (51). In mouse models, Bregs appear to be derived from either BII (follicular) or Marginal Zone (MZ) cells. A CD1d⁺ CD19⁺⁺ regulatory B cell subset has been identified in mice which strongly resembles MZ cells and protects against colitis, through interactions with Tregs (52). These observations not only demonstrate that these cells play a crucial pathogenic role in health and disease, but also that B cells can contribute to immune outcomes in general via their polarised cytokine expression.

Studies in mouse models of skin cancer suggest that there are certain circumstances where B cells can exert a distinctly immunosuppressive pro-tumour effect (53). A subset of “tumour-evoked” Bregs has been shown to facilitate the metastasis of breast cancer tumours through the induction of Tregs via TGF β secretion (54). Importantly, tumour-associated antigens such as 5-lipoxygenase (breast cancer; (55) and placental growth factor (glioblastoma; (56) have been shown to induce IL-10-production in tumour-infiltrating B cells in murine models. It may therefore be possible that B cells in the tumour-microenvironment are coerced into developing a regulatory phenotype (TGF β ⁺ and IL-10⁺), and that tumour-derived antigens in general could induce B cell IL-10-expression (57). As expected, Bregs have been shown to directly contribute to tumour-progression as a result of their anti-inflammatory properties (58).

At present there is a lack of substantial information relating to the role of immunosuppressive IL-10-producing B cells in affecting tumour progression in humans. Perhaps the most significant study showed that Granzyme-B-expressing IL-10⁺ B cells induced by T cells IL-21 secretion infiltrate solid tumours of breast, cervical and ovarian cancer patients and inhibit T cell responses (59). In addition, IL-10⁺ transitional B cells have been shown to be upregulated in gastric cancer patient

peripheral blood and tumours and are functionally capable of suppressing Th1 cytokine secretion by T cells and inducing Tregs (60).

The current evidence for regulatory B cells in melanoma tissue microenvironments rests upon a study demonstrating that adoptive transfer of B1a B cells (possessing an IL-10+ regulatory phenotype) into wild-type mice significantly exacerbated B16F10 melanoma growth, illustrating that IL-10-producing B cells can directly contribute to melanoma tumour-progression (61). A separate murine model of squamous cell carcinoma obtained similar conclusions (62). Finally, Bregs have been shown to suppress cutaneous inflammation in a mouse model of psoriasis-like inflammation (63). To date, there have been no investigations into the role of IL-10-producing B cells in patients with melanoma.

Tumour-infiltrating regulatory B cells therefore represent a potential mechanism of tumour-mediated immune escape that in certain conditions may outweigh antigen-presentation and anti-tumour antibody-mediated effector mechanisms to skew the overall impact of B cell infiltrates towards a neutral or even a negative effect, and mandates studies into the induction of regulatory B cells in melanoma patients and correlation with overall survival.

Influence of the Th2-biased melanoma microenvironment on pro-tumour properties of B cells

Consistent with the contributions of regulatory B cells, the microenvironments of melanoma solid tumours are characteristically considered as harbouring Th2-biased cytokine expression profiles (64), typical of chronic inflammatory conditions. These microenvironments typically confer pro-tumour properties including promoting angiogenesis and inhibiting cell-mediated responses, such as those mediated by cytotoxic T-lymphocytes (CTLs). The classic Th2 cytokine is interleukin-4 which, in addition to interleukin-13, may induce tumour clearance (65). However, IL-10 produced by Bregs, Tregs and M2-type macrophages in the tumour microenvironment can, in addition to its potent suppressive effects upon CTLs, trigger a modified or alternative Th2 response by inducing B cell IgG4 subclass switching in the presence of IL-4 (66).

Indeed, it has been shown *in vitro* that these Th2-biased microenvironments favour alternatively activated humoral immunity which confers a shift in B cell antibody expression towards IgG4 (67). Early evidence has accordingly demonstrated that IgG4 serum levels are dysregulated in melanoma patients, particularly in advanced disease settings (68). IgG4 subclass antibodies are structurally distinct to IgG1 antibodies and confer a weaker ability to bind and stimulate effector cells such as macrophages to initiate ADCP and ADCC through their Fc-gamma receptors (69). It has been shown that these antibodies compete with IgG1 for Fc-gamma receptor sites resulting in a reduced degree of immune cell activation and tumour-cell killing when secreted in large quantities (34,67). Recent evidence also suggests that Fc-Fc interactions between IgG4 and cancer-specific IgG1 antibodies may confer an important aspect of tumour immune evasion (70). Interestingly, the same study also demonstrated that the IgG4 subclass CPI antibody Nivolumab participated in Fc-interactions with IgG1 subclasses and promoted cancer growth in a murine model of colon cancer. The pro-tumour role of IgG4 subclasses may appropriately explain observations of possible hyperprogressive disease seen in some CPI patients.

IgG4 is therefore a negative prognostic indicator of patient overall survival, and these Th2 biased microenvironments offer a form of immune evasion which is mediated by IL-10 (71,72). Interestingly, recent evidence has also revealed that an IgG4-expressing CD49b⁺ CD73⁺ B cell subset expressing proangiogenic cytokines including VEGF is upregulated in the circulation of melanoma patients, highlighting an additional and previously unknown mechanism by which tumour-infiltrating B cells may contribute to tumour progression (73).

The role of B cells in checkpoint inhibitor therapy responses

Checkpoint inhibitors are designed to regulate T cell effector functions, however there is growing interest in the contribution of B cell responses to patient outcomes. This has been examined in several recent studies, the findings are summarised in Table 1 and Figure 2.

Tumour-infiltrating B cell signatures in relation to CPI treatment response:

Further evidence supports the notion that specific B cell subsets may hold prognostic value in CPI treatment response in advanced melanoma. B cell receptor repertoire and B cell phenotype were analysed in 473 cutaneous melanoma specimens. The

absence of B cell immunoglobulin heavy chain γ (IGHG) gene was found to correlate with lack of response to anti-CTLA-4 but this was not shown in the context of anti-PD-1 therapy (22). In addition, a higher proportion of IL-10 secreting B cells was associated with lack of response to anti-CTLA-4, but was not correlated with response to anti-PD-1 therapy (22). A phase 2 trial of neoadjuvant CPI (neoadjuvant Nivolumab (n=11) *versus* neoadjuvant Nivolumab plus Ipilimumab (n=12)) (74) also highlighted the importance of B cells in CPI response. While baseline tumour-infiltrating B cell counts did not differ significantly between responders and non-responders in this small sample size, early on-treatment tumour samples showed increased tumour-infiltrating B cell frequency (as defined by CD20 immunostaining) in treatment responders compared with non-responders. Helmink et al. used the same cohort and performed bulk RNA sequencing from baseline and early on-treatment tumour biopsy samples (75) and found that B cell markers were the most differentially expressed genes in responders to therapy compared with non-responders. They identified significantly increased clonal counts for both immunoglobulin heavy and light chains and increased BCR diversity in responders than in non-responders, which suggests an active role for B cells in anti-tumour immunity. Pathways upregulated in responders as compared to non-responders included those consistent with increased immune activity such as CXCR4 signalling, cytokine receptor interaction and chemokine signalling pathways. Phenotyping of B cells in tumours identified significantly higher frequency of memory B cells in responders *versus* non-responders at baseline and early on-treatment. They corroborated their findings in a further cohort of 18 patients with melanoma treated with adjuvant or neoadjuvant CPI. Unlike tumour-derived B cells, a peripheral blood B cell phenotype was not found to correlate with treatment outcome (75). Since most samples in this study were blood or metastatic lymph node deposits, the cohort was relatively small and the samples were sourced either at baseline or early on-treatment, these findings would benefit from evaluations of larger and more comprehensive studies.

Furthermore, in a cohort of 39 patients with advanced melanoma, early changes were observed in peripheral B cell subsets upon one cycle of combination CPI therapy (Ipilimumab and Nivolumab): particularly a significant decrease in circulating B cells in patients who underwent combination CPI as opposed to those treated with a monotherapy anti-CTLA-4 or anti-PD-1 (76). No significant differences were found between naïve and memory B cell subpopulations. Despite the overall decline in B cell numbers, a marked increase was observed in the density of: class-switched

memory B cells, plasmablasts and CD21^{lo} B cell sub-populations (76). Of note, significant changes in the latter were only exhibited in patients who underwent combination or anti-CTLA-4 treatment. This is consistent with findings from a previous study whereby, loss of circulating B cells and increase in CD21^{lo} B cells was reported in patients treated with an anti-CTLA-4 agent (77). Detailed evaluation of the CD21^{lo} B cell compartment revealed equal concentrations of naïve and memory B cells at baseline, but moderate increase in memory B cells after combination checkpoint inhibition. Furthermore, B cell receptor sequencing demonstrated that CD21^{lo} B cells possessed higher: clonality, maximal clone frequency, and SHM frequency. Therefore, CD21^{lo} B cells were shown as a phenotypically distinct B cell subset present in melanoma patients, which increased in frequency following combination checkpoint inhibition, and suggest an active B cell response in the TME. PD-1 expression on the CD21^{lo} B cells was higher than on other B cell subsets, suggesting that this population may be specifically modulated by anti-PD-1 therapy. This study failed to find an association between B cell changes and clinical response. The extrapolation of these findings may be limited due to the small sample size and the fact that on-treatment peripheral blood samples were only taken at an early time point after commencement of CPI therefore may not be representative of the full response.

CPI-associated modulation of plasmablasts:

There is some evidence that plasmablasts (PB) may be directly modulated by CPI. Circulating plasmablast levels were significantly enhanced in patients undergoing treatment with anti-CTLA-4 immunotherapy (78). When immunoglobulins produced by the PBs were sequenced, there was evidence of clonal expansion, SMH and class switching, indicative of an active B cell response (78). Immunostaining of 41 metastatic melanoma specimens revealed that a plasmablast-like phenotype (CD19+CD20+CD38+CD138-) is the most frequently observed B cell subset in the TME (41). Furthermore, circulating B cells could be induced to differentiate into plasmablast-like cells when exposed to autologous melanoma secretomes *in vitro* (41). In this study tumour-associated B-cells also expressed T-cell and macrophage chemoattractant mediators (CCL3, CCL3L1, CCL4, CCL5, CCL28, and CXCL16) suggesting that B-cells can orchestrate the immune response in the TME by modulating other cell populations. Increased expression of immunosuppressive cytokines IL-10 and TGFβ1 was reported in plasmablast-like and memory B cell subpopulations, consistent with a potential regulatory role. However, these signatures may also be associated with treatment response: proteomics and RNA-

seq data were used to define a tumour-induced plasmablast-like population (TIBP) functional signature and this signature was evaluated in a historical dataset from patients treated with Nivolumab. In this analysis, the presence of this TIBP signature at baseline correlated with better response to anti-PD-1 treatment (41).

A role for tertiary lymphoid structures:

The importance of TLS in response to CPI has also been investigated. Helmink et al. delved deeper into the localisation of B cells within their original cohort of 23 tumours and identified a higher density of CD20+ B cells in TLS and a higher TLS:tumour area ratio in early on-treatment samples from responders *versus* biopsies from non-responders (75). They also illustrated that CD20 B cells often co-localised with CD4, CD8 and FOXP3 T cells. A second study also identified CD20 B cells in 25% of 177 melanoma specimens (27). CD20 B cell clusters were surrounded by CD4 T cells. Increased expression of known molecular markers of TLS (CXCL13, CXCR5 and DC-LAMP) was described and immunostaining confirmed the co-localisation of CXCL13 and CXCR5 with the CD20 B cell marker. A TLS gene signature in melanoma was defined and integrated into four historical melanoma cohorts treated with CPI. This TLS gene signature at baseline was associated with significantly-increased survival with anti-CTLA-4 therapy and this was independent of the tumour mutational burden which may be a predictor for TLS formation (27). The role of B cells in TLS in response to CPI has also been highlighted in a third study in the treatment of sarcoma (79). In this, 213 soft tissue sarcomas were stratified according to the composition of the TME. Those with highest density of B cells and TLSs were associated with the highest response rate to CPI. The study identified several immune cell-related gene signatures and the only one significantly associated with extended overall survival was the B cell signature (79).

Taken together these studies highlight that B cell phenotype, localization with TLS and B cell gene expression may be directly modulated by CPI and could be relevant to treatment outcomes. The differential results seen with different agents and combination therapy likely reflect the distinct mechanisms of action of each treatment (80).

Antibody responses in melanoma

A key feature of B cells is their ability to produce antibodies. B cells in the tumour can undergo class switching and affinity maturation, producing tumour-specific or non-specific antibodies of different isotypes (24) and could exert anti-tumour or pro-tumour roles (81). Human antibodies are tetrameric glycoproteins formed by two heavy and two light polypeptide chains. Heavy and light chains assemble in a Y shaped structure, defined by a Fab region (formed by two identical Fab fragments, composed by the light chain and part of the heavy chain), an Fc region (formed by the constant portion of the two heavy chains) and a hinge region (joining Fab and Fc regions). The variable regions of heavy and light chains are assembled to form two identical antigen binding sites. The heavy chain constant region determines the isotype of the antibody, with a characteristic hinge and Fc region. The hinge region is responsible for the flexibility of the Fab arms which can affect both antigen binding and effector function and for some isotypes, such as IgG4, these can be involved in a process called Fab Arm Exchange, which results in bispecific (IgG4) antibodies (72). The Fc region is involved in the binding to Fc receptors (FcRs) and C1q (complement) and is responsible for antibody mediated effector functions such as ADCC, ADCP and CDC (80).

Depending on their isotype and specificity, antibodies can have opposite roles in the tumour associated immune response. Different antibody isotypes have different affinity for FcRs and C1q and consequently have different ability to trigger effector functions. Tumour specific antibodies with high affinity for FcRs and C1q (such as IgG1) have the potential to engender anti-tumour effects, mediating ADCC, ADCP, CDC or to support antigen presentation by mediating the uptake of tumour antigens by APCs such as macrophages and dendritic cells (82). Furthermore, membrane bound tumour-specific antibodies on the surface of B cells can engage with tumour cells and trigger B cell mediated cytotoxicity. Evidence of B cell mediated cytotoxicity via granzyme B and TRAIL has been shown in hepatocellular carcinoma (83). IgG class antibodies have been found to be tumour-specific, and this has been reported in melanoma (84) and other cancer types and has been correlated with a favourable prognosis (85–88).

Tumour associated antibodies can also have either pro-tumour effects or their anti-tumour effects may be regulated by tumours. Tumour-specific antibody-mediated immune cell activation and anti-tumour cytotoxicity may also be impaired by expression of inhibitory Fc receptors (FcγRIIb) in the TME and downstream immunoreceptor tyrosine-based inhibitory motif (ITIM) domain signalling (89,90).

Tumour-specific antibodies may be expressed as isotypes (such as IgG2 or IgG4) with poor affinity for activating FcRs or complement. These isotypes can be ineffective in triggering anti-tumour responses like ADCC, ADCP, CDC and antigen presentation (72). Furthermore, IgG4 antibodies can engage the inhibitory receptor FcγRIIb. This might further impair anti-tumour responses, blocking IgG1 mediated effector function (67). Tumour associated antibodies not specific for the tumour can also serve as a decoy blocking other potentially immuno-activating and tumour-reactive antibodies from engaging with immune effector cells. This can contribute to pro-tumour effects such as ineffective T cell mediated responses or B effector functions. It is also possible that tumour-specific antibodies accumulating at the tumour site over time forming immune-complexes, promoting chronic inflammation and immunosuppressive myeloid cell phenotypes (82).

Isotypes like IgG1 and IgG3 have high capacity to trigger ADCC and facilitate complement activation. The IgG1 isotype is usually associated with anti-tumour responses while TCGA RNA-seq data analysis of melanoma samples showed that IgG3 is either neutral or is associated with a negative prognosis. This could be explained by the fact that IgG3 has a shorter half-life (1 week) compared to other isotypes (IgG1, 2 weeks). Furthermore B cells expressing IgG3 isotype antibodies usually undergo less antibody SHM and therefore IgG3 antibodies may have lower affinity for the target antigen whilst still able to occupy FcRs (91). IgG2 has poor ability to bind FcRs and C1q. Evidence of antibody responses switched to IgG2 has been found in melanoma (26). IgG4 has poor ability to bind activating FcRs whilst able to bind the inhibitory receptor FcγRIIb and has complement-activating ability. Evidence of IgG4 antibody responses have been found in the melanoma TME and serum IgG4 has been associated with poor prognosis in melanoma (36,67,71).

TCGA RNA-seq data analyses also suggest that IgD, IgE and IgA antibodies may be associated with poor prognosis in melanoma (91). One possible explanation is that IgD can bind basophils resulting in the production of cytokines like IL-4, IL-5, IL-13, BAFF and APRIL which may support class switching to IgA and IgE and a Th2 immune response. Furthermore, the presence of IgE antibodies is usually associated with IL-4, which can further support a Th2 immune response. IgA antibodies have been found in melanoma and are associated with poor prognosis. This could be explained by the immune-suppressive phenotype of IgA⁺ B cells, expressing IL-10 and PD-L1 and by the ability of IgA⁺ B cells to support the expansion of T regulatory (Treg) cells which in turn secrete TGFβ supporting IgA class switch. Melanoma-

associated IgA have been found to correlate with poor clonality, suggesting that, in the tumour, class switching to IgA is a consequence of inflammation and not of an antigen-driven response, and that tumour associated IgA could be non-tumour specific (82).

Associations of B cell expressed antibodies with CPI treatment

Based on mounting evidence in support of potential roles of various antibody isotypes in melanoma, it is not surprising that antibody responses have been studied and are thought to correlate with outcome of CPI therapy. In the context of B cells in HIV infection, blocking of PD-1 has been shown to increase viral antigen-specific antibody responses (92). This may lend merit to the notion that PD-1-specific CPI could improve tumour-specific antibody responses (93).

One study investigated whether IgG antibody and isotype levels correlated with anti-tumour response and survival following CPI therapy. IgG subclass analysis was performed on serum samples at baseline from 49 patients with melanoma treated with anti-PD1 antibody (Nivolumab or Pembrolizumab (86%), Nivolumab plus Ipilimumab (10%), or Ipilimumab (4%). A positive correlation with progression-free survival (PFS) was found for high titres of total IgG, IgG1, IgG2 and IgG3, while a positive correlation with overall survival (OS) was found to be significant only for the IgG2 subclass (94). This result is quite interesting since IgG2 is usually not associated with anti-tumour immune responses due to its poor ability to trigger ADCC, ADCP, CDC and facilitate antigen presentation, but may reflect activation of a tumour antigen-specific response triggered by CPI.

A different preliminary study compared the antibody sequence repertoires of 26 patients with melanoma treated with Pembrolizumab (9), Ipilimumab (8) or Nivolumab and Ipilimumab combination (9). It was found that long-term responders had higher levels of SHM after initiation of treatment, and on-treatment compared to the non-responders. Comparison of lineages between patients identified antibodies with high sequence similarity suggesting these antibodies may have arisen from convergent selection, *i.e.*, different patients raising antibodies against shared or similar epitopes. Moreover, when antibody lineages were analysed and compared among patients, antibodies with high sequence similarity were found, suggesting these antibodies may have arisen from convergent selection and that the patients

may be producing antibodies against shared epitopes. Of note, IgG2 was the most frequent isotype of these antibodies. Furthermore IgG2 was higher in responders compared to non-responders (95).

In a different study, melanoma patients treated with CPI (monotherapy with Nivolumab, Pembrolizumab or Ipilimumab, or the combination of Nivolumab and Ipilimumab), the responder group had higher titres of antibodies specific for the tumour associated antigens MDA (TRP1/TYRP1, TRP2/TYRP2, gp100, MelanA/MART1) and the Cancer-Testis antigen NY-ESO-1 at baseline, compared to non-responders. Serum analyses showed that NY-ESO-1, TRP1/TYRP1 and TRP2/TYRP2 specific antibodies consisted of several IgG subclasses while the IgG antibodies for MelanA were mostly IgG1 and the antibodies for gp100 were of the IgG2 isotype (84). Interestingly, none of these antibodies were IgG4, supporting the fact that IgG4 antibodies are not associated with a good prognosis in melanoma patients (67,71,84,94).

In patients with metastatic melanoma who were treated with anti-CTLA-4 combined with anti-angiogenic VEGF-A-targeted treatment, there was an increase of antibody titres recognising the immunoregulatory protein Galectin-1 (Gal-1). Anti-Gal-1 antibody titres in turn correlated with better disease outcome. Higher frequencies of complete or partial responses and improved overall survival were seen in these individuals. Gal-1 has been reported to be upregulated in many tumour types including melanoma and is usually associated with poorer survival, contributing to tumour growth, angiogenesis, metastasis and immune evasion (96).

Taken together, these results suggest that checkpoint blockade-associated restoration of T cell activity may also contribute to increasing the effectiveness of the humoral response and highlights the importance of the expressed antibodies as part of the anti-tumour activity. Furthermore, these studies suggest that melanoma-specific antibodies in pre-treatment sera may be promising indicators of CPI immunotherapy response and require further study.

Correlations with immune related adverse events in CPI therapy

Activation of immune responses as a result of checkpoint inhibition can lead to compromised immune self-tolerance and can consequently result in the development

of irAEs, toxicities that mimic autoimmunity. The nature and frequency of irAEs varies amongst CPI agents and can include effects on the gastrointestinal tract (colitis), liver (hepatitis), lung (pneumonitis) and endocrine systems, including hypophysitis and insulin-dependent diabetes. The effects of anti-CTLA4 associated irAEs are generally more severe than those from anti PD-1 inhibition. Whether irAEs represent *de novo* events or whether they represent an unmasking of underlying immune mediated disease remains unclear (6).

Modulation of B cell phenotype has been correlated with irAE in patients undergoing CPI therapy. A recent study (described above) demonstrated an overall decline in B cell numbers but increased plasmablasts and CD21^{lo} B cell subsets in patients given combination checkpoint inhibition (Table 1) (76). These CPI induced B cell changes positively correlated with frequency and severity of irAEs. Furthermore, patients with altered B cell populations were more likely to develop multi-organ immunotoxicity than those without, with these changes preceding toxicity by a median of 3 weeks.

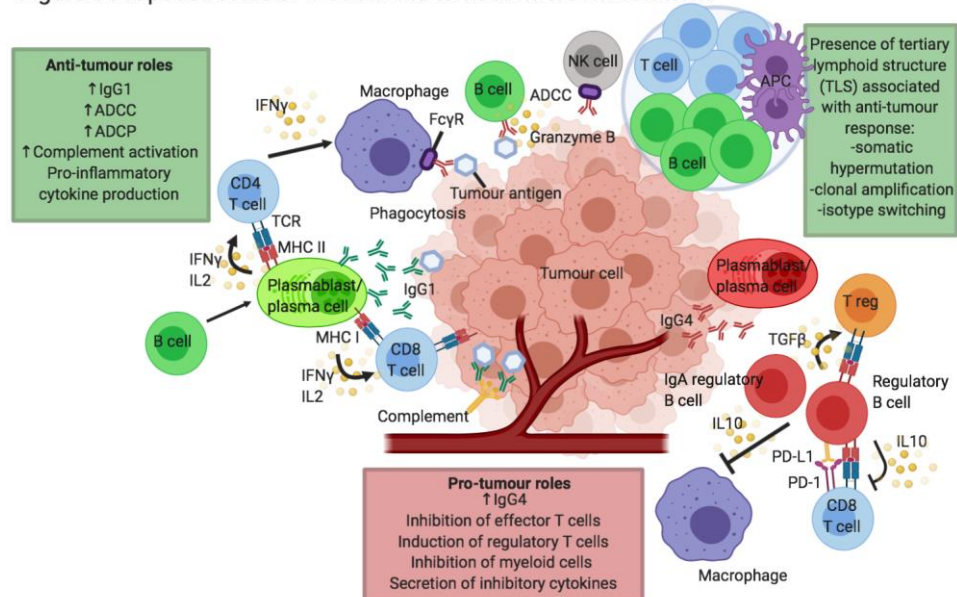
B cells are proposed to contribute to autoimmunity via multiple mechanisms including as antibody secreting cells, cytokine producers, APCs and immunoregulators. Given the important role of the checkpoint pathways in regulating immune homeostasis, it is not surprising that activation of the CTLA-4 pathway has been extensively studied in the treatment of autoimmunity. Specifically Abatecept, a fusion protein of CTLA-4 and IgG1 Fc portion (CTLA-4-Ig), is approved for abrogating immune overactivity in rheumatoid arthritis (97–99). CTLA-4-Ig is able to directly bind CD80/86; therefore, preventing CD28 binding and T cell co-stimulation. B cells are known to express CD80/86 and have been shown in mouse models and *in vitro* to be a direct target for CTLA-Ig. In synovial biopsy samples, CTLA-4-Ig has been shown to decrease B cell infiltrates (97). Humoral responses to T cell dependent and T cell independent antigenic stimulation are also suppressed by CTLA-4-Ig *in vivo* (100,101). These insights from autoimmunity may also have relevance to our understanding of how CPI modification of B cells may induce irAE.

The presence of autoantibodies in CPI induced irAE has been detected in a similar fashion to those seen in autoimmune disease in the absence CPI treatment. For example some patients suffering with CPI-induced hypothyroidism have raised titres of anti-thyroperoxidase antibody and/or anti-thyrotropin receptor antibodies (102,103). However, as these autoantibodies are not universally detected in CPI thyroid disorders it is unclear whether they play a causative role. Similar observations

have been made in a range of CPI induced autoimmune phenomena including cases of myasthenia gravis (104), type 1 diabetes mellitus (105) and autoimmune haemolytic anaemia (106). One study analysed 23 common autoantibodies in baseline peripheral blood and post CPI treatment in 133 melanoma patients treated with Ipilimumab (107). This showed that 19% of patients developed autoantibodies on-treatment and a trend of association with onset of irAE which did not reach statistical significance. In this study, organ-specific autoantibodies did not correlate with irAE.

CPI induced irAE present a clinical challenge for patient selection and can lead to treatment discontinuation. Insights into the complex balance between activation and suppression of B cell subsets may infer consequences for both treatment response and irAE. In addition, the potential contributions of autoantibodies to irAE warrant further investigation and may be of direct relevance to B cell biology in the context of CPI.

Figure 1 Proposed roles of B cell in the tumour microenvironment



Harnessing the humoral immune response in melanoma immunotherapy

Data from several studies suggest that B cells can play multi-faceted roles in the immune response to melanoma and that melanoma-associated immune evasion mechanisms involve modulating B cell immunity in addition to suppressing T cell activation. Despite the impressive efficacy demonstrated by CPI, a large proportion of patients fail to respond and/or develop irAEs (4,103-104). Modulated B cell and

antibody repertoires could either obstruct or facilitate the efficacy of CPI, depending on an array of factors in the TME. Further research identifying specific cellular surface markers and methods to clearly dissect the various B cell phenotypes will help improve our understanding of B cell immunity in melanoma and whether and how this is altered by CPI. This may then be exploited in clinical trials to help identify therapeutic strategies which can be combined with CPI to improve response. This may include the targeting of B cells; a pilot study using Rituximab (anti-CD20) to deplete B cells in melanoma patients increased the median time without recurrence from 6 to 42 months (110). On the other hand, B cell depletion therapy may have contrasting effects dependent on the stage of melanoma in mouse models (41). Given the newfound appreciation of the complexity of the TIL-B subpopulations, a more effective approach may involve depleting specific pro-tumour subsets, such as Bregs and IL-10+ B cells, whilst promoting anti-tumour subsets which could be combined with CPI as an adjuvant (24). PD1-PD-L1 engagement plays a role in suppressing B cell-mediated T cell activation, influencing CPI efficacy. Inhibition of PD-1 expressed on B cells may also be a mechanism through which CPI directly modulate B cell responses.

The targeting of the immunosuppressive TME cytokines IL-10, VEGF or TGF- β may effectively reinstate robust anti-tumour immunity by preventing IgG1 to IgG4 class switching and promoting anti-tumour B cell subsets to engage in tumour rejection. The hostile TME may be taken into consideration in the engineering of therapeutic antibody candidates, with the aim of potentially producing antibodies more resilient to immune suppression. CPI antibodies could additionally be engineered to have reduced binding to inhibitory Fc receptors, increased binding to activatory Fc receptors, or consider alternative isotypes such as IgE which has been implicated to improve immune surveillance in melanoma (111–113).

Aside from clinical response, irAEs are a major challenge of CPI in melanoma, as 96% of patients receiving combined CPI experienced at least one irAE in the CheckMate067 trial (4). Immune monitoring for early alterations in B cells following CPI may hold some promise in determining those at risk of developing irAEs. If confirmed to contribute to irAEs, therapies targeting plasmablasts and CD21^{lo} B cells could be explored to reduce the risk. Serum and melanoma IgG4 and IgG4+ B cell levels may also be monitored and therapies targeting these subsets could be conducted prior to CPI to establish a 'hot' tumour more likely to respond to CPI. Throughout CPI, B cell and antibody levels may be monitored to determine

modulation and predict prognostic outcomes and spikes may be countered through further depletion methods. Similar strategies could be conducted with immunosuppressive cytokines in the TME, anti-tumour B cell subsets and IgG1 levels.

Conclusion

It is increasingly appreciated that melanoma can develop intricate mechanisms to modulate the humoral immune response and B cell immunity may also play a significant role in the efficacy and safety of CPI in melanoma. Taking all into consideration, it is highly conceivable that the promotion of anti-tumour B cell immunity with targeted therapies can also have clinical relevance in CPI response. Furthermore, immunomonitoring of humoral responses prior to, during and following CPI in predicting irAEs should be explored. Given the complexity of tumour:immune cross-talk, utilising B cells and antibodies as prognostic/predictive biomarkers, in patient stratification and in immunomonitoring remains largely unexplored. Factoring in B cell immunity may harbour significant potential to overcome the two greatest challenges associated with CPI in melanoma: response and irAEs. Further studies in patients, *in vivo* and *ex vivo* models of immuno-oncology, alongside clinical testing, are required to fully comprehend and exploit likely multifaceted contributions.

References:

1. Robbins HA, Clarke CA, Arron ST, Tatalovich Z, Kahn AR, Hernandez BY, Paddock L, Yanik EL, Lynch CF, Kasiske BL, et al. Melanoma Risk and Survival among Organ Transplant Recipients. *J Invest Dermatol* (2015) **135**:2657–2665. doi:10.1038/jid.2015.312
2. Shiels MS, Copeland G, Goodman MT, Harrell J, Lynch CF, Pawlish K, Pfeiffer RM, Engels EA. Cancer stage at diagnosis in patients infected with the human immunodeficiency virus and transplant recipients. *Cancer* (2015) **121**:2063–2071. doi:10.1002/cncr.29324
3. Larkin J, Chiarion-Sileni V, Gonzalez R, Grob J-J, Rutkowski P, Lao CD, Cowey CL, Schadendorf D, Wagstaff J, Dummer R, et al. Five-Year Survival with Combined Nivolumab and Ipilimumab in Advanced Melanoma. *N Engl J Med* (2019) **381**:1535–1546. doi:10.1056/NEJMoa1910836
4. Hodi FS, Chiarion-Sileni V, Gonzalez R, Grob J-J, Rutkowski P, Cowey CL, Lao CD, Schadendorf D, Wagstaff J, Dummer R, et al. Nivolumab plus ipilimumab or nivolumab alone versus ipilimumab alone in advanced melanoma (CheckMate 067): 4-year outcomes of a multicentre, randomised, phase 3 trial. *Lancet Oncol* (2018) **19**:1480–1492. doi:10.1016/S1473-2045(18)30700-9
5. FDA. <https://www.fda.gov/drugs/resources-information-approved-drugs/fda-approves-atezolizumab-braf-v600-unresectable-or-metastatic-melanoma>.
6. Ramos-Casals M, Brahmer JR, Callahan MK, Flores-Chávez A, Keegan N, Khamashta MA, Lambotte O, Mariette X, Prat A, Suárez-Almazor ME. Immune-related adverse events of checkpoint inhibitors. *Nat Rev Dis Prim* (2020) **6**: doi:10.1038/s41572-020-0160-6
7. Alegre ML, Noel PJ, Eisfelder BJ, Chuang E, Clark MR, Reiner SL, Thompson CB. Regulation of surface and intracellular expression of CTLA4 on mouse T cells. *J Immunol* (1996) **157**:4762–4770.
8. Takahashi T, Tagami T, Yamazaki S, Uede T, Shimizu J, Sakaguchi N, Mak TW, Sakaguchi S. Immunologic self-tolerance maintained by CD25(+)CD4(+) regulatory T cells constitutively expressing cytotoxic T lymphocyte-associated antigen 4. *J Exp Med* (2000) **192**:303–310. doi:10.1084/jem.192.2.303
9. Walunas TL, Lenschow DJ, Bakker CY, Linsley PS, Freeman GJ, Green JM, Thompson CB, Bluestone JA. CTLA-4 can function as a negative regulator of T cell activation. *Immunity* (1994) **1**:405–413. doi:10.1016/1074-7613(94)90071-x
10. Sansom DM. CD28, CTLA-4 and their ligands: who does what and to whom? *Immunology* (2000) **101**:169–177. doi:10.1046/j.1365-2567.2000.00121.x
11. Keir ME, Butte MJ, Freeman GJ, Sharpe AH. PD-1 and its ligands in tolerance and immunity. *Annu Rev Immunol* (2008) **26**:677–704. doi:10.1146/annurev.immunol.26.021607.090331
12. Agata Y, Kawasaki A, Nishimura H, Ishida Y, Tsubata T, Yagita H, Honjo T. Expression of the PD-1 antigen on the surface of stimulated mouse T and B lymphocytes. *Int Immunol* (1996) **8**:765–772. doi:10.1093/intimm/8.5.765
13. Yearley JH, Gibson C, Yu N, Moon C, Murphy E, Juco J, Lunceford J, Cheng J, Chow LQM, Seiwert TY, et al. PD-L2 Expression in Human Tumors: Relevance to Anti-PD-1 Therapy in Cancer. *Clin cancer Res an Off J Am Assoc Cancer Res* (2017) **23**:3158–3167. doi:10.1158/1078-0432.CCR-16-1761
14. Hui E, Cheung J, Zhu J, Su X, Taylor MJ, Wallweber HA, Sasmal DK, Huang J, Kim JM, Mellman I, et al. T cell costimulatory receptor CD28 is a primary target for PD-1-mediated inhibition. *Science* (2017) **355**:1428–1433. doi:10.1126/science.aaf1292

15. Balkwill FR, Capasso M, Hagemann T. The tumor microenvironment at a glance. *J Cell Sci* (2012) **125**:5591–5596. doi:10.1242/jcs.116392
16. Yokosuka T, Takamatsu M, Kobayashi-Imanishi W, Hashimoto-Tane A, Azuma M, Saito T. Programmed cell death 1 forms negative costimulatory microclusters that directly inhibit T cell receptor signaling by recruiting phosphatase SHP2. *J Exp Med* (2012) **209**:1201–1217. doi:10.1084/jem.20112741
17. Walker LSK, Sansom DM. Confusing signals: recent progress in CTLA-4 biology. *Trends Immunol* (2015) **36**:63–70. doi:10.1016/j.it.2014.12.001
18. Qureshi OS, Zheng Y, Nakamura K, Attridge K, Manzotti C, Schmidt EM, Baker J, Jeffery LE, Kaur S, Briggs Z, et al. Trans-endocytosis of CD80 and CD86: a molecular basis for the cell-extrinsic function of CTLA-4. *Science* (2011) **332**:600–603. doi:10.1126/science.1202947
19. Schumacher TN, Schreiber RD. Neoantigens in cancer immunotherapy. *Science* (80-) (2015) **348**:69–74. doi:10.1126/science.aaa4971
20. Fu Q, Chen N, Ge C, Li R, Li Z, Zeng B, Li C, Wang Y, Xue Y, Song X, et al. Prognostic value of tumor-infiltrating lymphocytes in melanoma: a systematic review and meta-analysis. *Oncoimmunology* (2019) **8**:1593806. doi:10.1080/2162402X.2019.1593806
21. Fridman WH, Pagès F, Sautès-Fridman C, Galon J. The immune contexture in human tumours: impact on clinical outcome. *Nat Rev Cancer* (2012) **12**:298–306. doi:10.1038/nrc3245
22. Selitsky SR, Mose LE, Smith CC, Chai S, Hoadley KA, Dittmer DP, Moschos SJ, Parker JS, Vincent BG. Prognostic value of B cells in cutaneous melanoma. *Genome Med* (2019) **11**:1–11. doi:10.1186/s13073-019-0647-5
23. Cursons J, Souza-Fonseca-Guimaraes F, Foroutan M, Anderson A, Holland F, Hediye-Zadeh S, Behren A, Huntington ND, Davis MJ. A Gene Signature Predicting Natural Killer Cell Infiltration and Improved Survival in Melanoma Patients. *Cancer Immunol Res* (2019) **7**:1162–1174. doi:10.1158/2326-6066.CIR-18-0500
24. Chiaruttini G, Mele S, Opzoomer J, Crescioli S, Ilieva KM, Lacy KE, Karagiannis SN. B cells and the humoral response in melanoma: The overlooked players of the tumor microenvironment. *Oncoimmunology* (2017) **6**:1–11. doi:10.1080/2162402X.2017.1294296
25. Germain C, Gnjjatic S, Dieu-Nosjean M-C. Tertiary Lymphoid Structure-Associated B Cells are Key Players in Anti-Tumor Immunity. *Front Immunol* (2015) **6**:67. doi:10.3389/fimmu.2015.00067
26. Cipponi A, Mercier M, Seremet T, Baurain J-F, Theate I, van den Oord J, Stas M, Boon T, Coulie PG, van Baren N. Neogenesis of lymphoid structures and antibody responses occur in human melanoma metastases. *Cancer Res* (2012) **72**:3997–4007. doi:10.1158/0008-5472.CAN-12-1377
27. Cabrita R, Lauss M, Sanna A, Donia M, Skaarup Larsen M, Mitra S, Johansson I, Phung B, Harbst K, Vallon-Christersson J, et al. Tertiary lymphoid structures improve immunotherapy and survival in melanoma. *Nature* (2020) doi:10.1038/s41586-019-1914-8
28. Munoz-Erazo L, Rhodes JL, Marion VC, Kemp RA. Tertiary lymphoid structures in cancer – considerations for patient prognosis. *Cell Mol Immunol* (2020) **17**:570–575. doi:10.1038/s41423-020-0457-0
29. Helmink BA. B cells and tertiary lymphoid structures promote immunotherapy response. *Nature* (2020) **577**: doi:10.1038/s41586-019-1922-8
30. Ladányi A. Prognostic and predictive significance of immune cells infiltrating cutaneous melanoma. *Pigment Cell Melanoma Res* (2015) **28**:490–500. doi:10.1111/pcmr.12371
31. Hussein MR, Elasers DAH, Fadel SA, Omar AEM. Immunohistological characterisation of tumour infiltrating lymphocytes in melanocytic skin lesions.

- J Clin Pathol* (2006) **59**:316–324. doi:10.1136/jcp.2005.028860
32. Hillen F, Baeten CIM, Van De Winkel A, Creyten D, Van Der Schaft DWJ, Winnepeninckx V, Griffioen AW. Leukocyte infiltration and tumor cell plasticity are parameters of aggressiveness in primary cutaneous melanoma. *Cancer Immunol Immunother* (2008) **57**:97–106. doi:10.1007/s00262-007-0353-9
 33. Martinez-Rodriguez M, Thompson AK, Monteagudo C. A significant percentage of CD20-positive TILs correlates with poor prognosis in patients with primary cutaneous malignant melanoma. *Histopathology* (2014) **65**:726–728. doi:10.1111/his.12437
 34. Karagiannis P, Gilbert AE, Nestle FO, Karagiannis SN. IgG4 antibodies and cancer-associated inflammation. *Oncoimmunology* (2013) **2**:e24889. doi:10.4161/onci.24889
 35. Saul L, Ilieva KM, Bax HJ, Karagiannis P, Correa I, Rodriguez-Hernandez I, Josephs DH, Tosi I, Egbuniwe IU, Lombardi S, et al. IgG subclass switching and clonal expansion in cutaneous melanoma and normal skin. *Sci Rep* (2016) **6**:1–12. doi:10.1038/srep29736
 36. Gilbert AE, Karagiannis P, Dodev T, Koers A, Lacy K, Josephs DH, Takhar P, Geh JLC, Healy C, Harries M, et al. Monitoring the systemic human memory B cell compartment of melanoma patients for anti-tumor IgG antibodies. *PLoS One* (2011) **6**:e19330. doi:10.1371/journal.pone.0019330
 37. Bosio FM, Wilmott JS, Volders N, Mercier M, Wouters J, Stas M, Blokx WAM, Massi D, Thompson JF, Scolyer RA, et al. Plasma cells in primary melanoma. Prognostic significance and possible role of IgA. *Mod Pathol* (2016) **29**:347–358. doi:10.1038/modpathol.2016.28
 38. Kim H, Hong SH, Kim JY, Kim IC, Park YW, Lee SJ, Song SW, Kim JJ, Park G, Kim TM, et al. Preclinical development of a humanized neutralizing antibody targeting HGF. *Exp Mol Med* (2017) **49**:e309–e309. doi:10.1038/emmm.2017.21
 39. Rodriguez-Pinto D. B cells as antigen presenting cells. *Cell Immunol* (2005) **238**:67–75. doi:10.1016/j.cellimm.2006.02.005
 40. Lund FE. Cytokine-producing B lymphocytes - key regulators of immunity. *Curr Opin Immunol* (2008) **20**:332–338. doi:10.1016/j.coi.2008.03.003
 41. Griss J, Bauer W, Wagner C, Simon M, Chen M, Grabmeier-Pfistershammer K, Maurer-Granofszky M, Roka F, Penz T, Bock C, et al. B cells sustain inflammation and predict response to immune checkpoint blockade in human melanoma. *Nat Commun* (2019) **10**: doi:10.1038/s41467-019-12160-2
 42. Endres R, Alimzhanov MB, Plitz T, Fütterer A, Kosco-Vilbois MH, Nedospasov SA, Rajewsky K, Pfeffer K. Mature follicular dendritic cell networks depend on expression of lymphotoxin beta receptor by radioresistant stromal cells and of lymphotoxin beta and tumor necrosis factor by B cells. *J Exp Med* (1999) **189**:159–168. doi:10.1084/jem.189.1.159
 43. Erdag G, Schaefer JT, Smolkin ME, Deacon DH, Shea SM, Dengel LT, Patterson JW, Slingluff CL. Immunotype and immunohistologic characteristics of tumor-infiltrating immune cells are associated with clinical outcome in metastatic melanoma. *Cancer Res* (2012) **72**:1070–1080. doi:10.1158/0008-5472.CAN-11-3218
 44. Garg K, Maurer M, Griss J, Brüggemann MC, Wolf IH, Wagner C, Willi N, Mertz KD, Wagner SN. Tumor-associated B cells in cutaneous primary melanoma and improved clinical outcome. *Hum Pathol* (2016) **54**:157–164. doi:10.1016/j.humpath.2016.03.022
 45. Blair PA, Noreña LY, Flores-Borja F, Rawlings DJ, Isenberg DA, Ehrenstein MR, Mauri C. CD19(+)CD24(hi)CD38(hi) B cells exhibit regulatory capacity in healthy individuals but are functionally impaired in systemic Lupus Erythematosus patients. *Immunity* (2010) **32**:129–140.

- doi:10.1016/j.immuni.2009.11.009
46. Ashour HM, Seif TM. The role of B cells in the induction of peripheral T cell tolerance. *J Leukoc Biol* (2007) **82**:1033–1039. doi:10.1189/jlb.0507310
 47. Mizoguchi A, Mizoguchi E, Takedatsu H, Blumberg RS, Bhan AK. Chronic intestinal inflammatory condition generates IL-10-producing regulatory B cell subset characterized by CD1d upregulation. *Immunity* (2002) **16**:219–230. doi:10.1016/S1074-7613(02)00274-1
 48. Duddy ME, Alter A, Bar-Or A. Distinct profiles of human B cell effector cytokines: a role in immune regulation? *J Immunol* (2004) **172**:3422–3427. doi:10.4049/jimmunol.172.6.3422
 49. Flores-Borja F, Bosma A, Ng D, Reddy V, Ehrenstein MR, Isenberg DA, Mauri C. CD19+CD24hiCD38hi B cells maintain regulatory T cells while limiting TH1 and TH17 differentiation. *Sci Transl Med* (2013) **5**:173ra23. doi:10.1126/scitranslmed.3005407
 50. Wu H, Su Z, Barnie PA. The role of B regulatory (B10) cells in inflammatory disorders and their potential as therapeutic targets. *Int Immunopharmacol* (2020) **78**: doi:10.1016/j.intimp.2019.106111
 51. Inoue S, Leitner WW, Golding B, Scott D. Inhibitory effects of B cells on antitumor immunity. *Cancer Res* (2006) **66**:7741–7747. doi:10.1158/0008-5472.CAN-05-3766
 52. Wei B, Velazquez P, Turovskaya O, Spricher K, Aranda R, Kronenberg M, Birnbaumer L, Braun J. Mesenteric B cells centrally inhibit CD4+ T cell colitis through interaction with regulatory T cell subsets. *Proc Natl Acad Sci U S A* (2005) **102**:2010–2015. doi:10.1073/pnas.0409449102
 53. Andreu P, Johansson M, Affara NI, Pucci F, Tan T, Junankar S, Korets L, Lam J, Tawfik D, DeNardo DG, et al. FcRgamma activation regulates inflammation-associated squamous carcinogenesis. *Cancer Cell* (2010) **17**:121–134. doi:10.1016/j.ccr.2009.12.019
 54. Olkhanud PB, Damdinsuren B, Bodogai M, Gress RE, Sen R, Wejksza K, Malchinkhuu E, Wersto RP, Biragyn A. Tumor-evoked regulatory B cells promote breast cancer metastasis by converting resting CD4+ T cells to T-regulatory cells. *Cancer Res* (2011) **71**:3505–3515. doi:10.1158/0008-5472.CAN-10-4316
 55. Wejksza K, Lee-Chang C, Bodogai M, Bonzo J, Gonzalez FJ, Lehrmann E, Becker K, Biragyn A. Cancer-Produced Metabolites of 5-Lipoxygenase Induce Tumor-Evoked Regulatory B Cells via Peroxisome Proliferator-Activated Receptor α . *J Immunol* (2013) **190**:2575–2584. doi:10.4049/jimmunol.1201920
 56. Han S, Feng S, Ren M, Ma E, Wang X, Xu L, Xu M. Glioma cell-derived placental growth factor induces regulatory B cells. *Int J Biochem Cell Biol* (2014) **57**:63–68. doi:10.1016/j.biocel.2014.10.005
 57. Zhang Y, Gallastegui N, Rosenblatt JD. Regulatory B cells in anti-tumor immunity. *Int Immunol* (2015) **27**:521–530. doi:10.1093/intimm/dxv034
 58. Horikawa M, Minard-Colin V, Matsushita T, Tedder TF. Regulatory B cell production of IL-10 inhibits lymphoma depletion during CD20 immunotherapy in mice. *J Clin Invest* (2011) **121**:4268–4280. doi:10.1172/JCI59266
 59. Lindner S, Dahlke K, Sontheimer K, Hagn M, Kaltenmeier C, Barth TFE, Beyer T, Reister F, Fabricius D, Lotfi R, et al. Interleukin 21-induced granzyme b-expressing b cells infiltrate tumors and regulate t cells. *Cancer Res* (2013) **73**:2468–2479. doi:10.1158/0008-5472.CAN-12-3450
 60. Wang W, Yuan X, Chen H, Xie G, Ma Y, Zheng Y, Zhou Y, Shen L. CD19+CD24hiCD38hi Bregs involved in downregulate helper T cells and upregulate regulatory T cells in gastric cancer. *Oncotarget* (2015) **6**:33486–33499. doi:10.18632/oncotarget.5588
 61. Kobayashi T, Oishi K, Okamura A, Maeda S, Komuro A, Hamaguchi Y, Fujimoto M, Takehara K, Matsushita T. Regulatory B1a Cells Suppress

- Melanoma Tumor Immunity via IL-10 Production and Inhibiting T Helper Type 1 Cytokine Production in Tumor-Infiltrating CD8+ T Cells. *J Invest Dermatol* (2019) **139**:1535-1544.e1. doi:10.1016/j.jid.2019.02.016
62. Schioppa T, Moore R, Thompson RG, Rosser EC, Kulbe H, Nedospasov S, Mauri C, Coussens LM, Balkwill FR. B regulatory cells and the tumor-promoting actions of TNF- α during squamous carcinogenesis. *Proc Natl Acad Sci U S A* (2011) **108**:10662–10667. doi:10.1073/pnas.1100994108
 63. Yanaba K, Kamata M, Ishiura N, Shibata S, Asano Y, Tada Y, Sugaya M, Kadono T, Tedder TF, Sato S. Regulatory B cells suppress imiquimod-induced, psoriasis-like skin inflammation. *J Leukoc Biol* (2013) **94**:563–573. doi:10.1189/jlb.1112562
 64. Ellyard JL, Simson L, Parish CR. Th2-mediated anti-tumour immunity: friend or foe? *Tissue Antigens* (2007) **70**:1–11. doi:10.1111/j.1399-0039.2007.00869.x
 65. Paul WE. Interleukin-4: a prototypic immunoregulatory lymphokine. *Blood* (1991) **77**:1859–1870.
 66. Satoguina JS, Weyand E, Larbi J, Hoerauf A. T Regulatory-1 Cells Induce IgG4 Production by B Cells: Role of IL-10. *J Immunol* (2005) **174**:4718–4726. doi:10.4049/jimmunol.174.8.4718
 67. Karagiannis P, Gilbert AE, Josephs DH, Ali N, Dodev T, Saul L, Correa I, Roberts L, Beddowes E, Koers A, et al. IgG4 subclass antibodies impair antitumor immunity in melanoma. *J Clin Invest* (2013) **123**:1457–1474. doi:10.1172/JCI65579
 68. Daveau M, Pavie-Fischer J, Rivat L, Rivat C, Ropartz C, Peter HH, Cesarini JP, Kourilsky FM. IgG4 subclass in malignant melanoma. *J Natl Cancer Inst* (1977) **58**:189–192. doi:10.1093/jnci/58.2.189
 69. Aalberse RC, Schuurman J. IgG4 breaking the rules. *Immunology* (2002) **105**:9–19. doi:10.1046/j.0019-2805.2001.01341.x
 70. Wang H, Xu Q, Zhao C, Zhu Z, Zhu X, Zhou J, Zhang S, Yang T, Zhang B, Li J, et al. An immune evasion mechanism with IgG4 playing an essential role in cancer and implication for immunotherapy. *J Immunother Cancer* (2020) **8**:e000661. doi:10.1136/jitc-2020-000661
 71. Karagiannis P, Villanova F, Josephs DH, Correa I, Van Hemelrijck M, Hobbs C, Saul L, Egbuniwe IU, Tosi I, Ilieva KM, et al. Elevated IgG4 in patient circulation is associated with the risk of disease progression in melanoma. *Oncoimmunology* (2015) **4**:e1032492–e1032492. doi:10.1080/2162402X.2015.1032492
 72. Crescioli S, Correa I, Karagiannis P, Davies AM, Sutton BJ, Nestle FO, Karagiannis SN. IgG4 Characteristics and Functions in Cancer Immunity. *Curr Allergy Asthma Rep* (2016) **16**:7. doi:10.1007/s11882-015-0580-7
 73. van de Veen W, Globinska A, Jansen K, Straumann A, Kubo T, Verschoor D, Wirz OF, Castro-Giner F, Tan G, Rückert B, et al. A novel proangiogenic B cell subset is increased in cancer and chronic inflammation. *Sci Adv* (2020) **6**:eaaz3559. doi:10.1126/sciadv.aaz3559
 74. Amaria RN, Reddy SM, Tawbi HA, Davies MA, Ross MI, Glitza IC, Cormier JN, Lewis C, Hwu WJ, Hanna E, et al. Neoadjuvant immune checkpoint blockade in high-risk resectable melanoma. *Nat Med* (2018) **24**:1649–1654. doi:10.1038/s41591-018-0197-1
 75. Helmink BA, Reddy SM, Gao J, Zhang S, Basar R, Thakur R, Yizhak K, Sade-Feldman M, Blando J, Han G, et al. B cells and tertiary lymphoid structures promote immunotherapy response. *Nature* (2020) doi:10.1038/s41586-019-1922-8
 76. Das R, Bar N, Ferreira M, Newman AM, Zhang L, Bailur JK, Bacchiocchi A, Kluger H, Wei W, Halaban R, et al. Early b cell changes predict autoimmunity following combination immune checkpoint blockade. *J Clin Invest* (2018) **128**:715–720. doi:10.1172/JCI96798

77. Kuehn HS, Ouyang W, Lo B, Deenick EK, Niemela JE, Avery DT, Schickel J-N, Tran DQ, Stoddard J, Zhang Y, et al. Immune dysregulation in human subjects with heterozygous germline mutations in CTLA4. *Science* (2014) **345**:1623–1627. doi:10.1126/science.1255904
78. DeFalco J, Harbell M, Manning-Bog A, Baia G, Scholz A, Millare B, Sumi M, Zhang D, Chu F, Dowd C, et al. Non-progressing cancer patients have persistent B cell responses expressing shared antibody paratopes that target public tumor antigens. *Clin Immunol* (2018) **187**:37–45. doi:10.1016/j.clim.2017.10.002
79. Petitprez F, de Reyniès A, Keung EZ, Chen TW-W, Sun C-M, Calderaro J, Jeng Y-M, Hsiao L-P, Lacroix L, Bougouïn A, et al. B cells are associated with survival and immunotherapy response in sarcoma. *Nature* (2020) **577**: doi:10.1038/s41586-019-1906-8
80. Scott AM, Wolchok JD, Old LJ. Antibody therapy of cancer. *Nat Rev Cancer* (2012) **12**:278–287. doi:10.1038/nrc3236
81. Sharonov G V, Serebrovskaya EO, Yuzhakova D V, Britanova O V, Chudakov DM. B cells, plasma cells and antibody repertoires in the tumour microenvironment. *Nat Rev Immunol* (2020) **20**:294–307. doi:10.1038/s41577-019-0257-x
82. Sharonov G V, Serebrovskaya EO, Yuzhakova D V. B cells, plasma cells and antibody repertoires in the tumour microenvironment. *Nat Rev Immunol* **1**: doi:10.1038/s41577-019-0257-x
83. Shi J-Y, Gao Q, Wang Z-C, Zhou J, Wang X-Y, Min Z-H, Shi Y-H, Shi G-M, Ding Z-B, Ke A-W, et al. Margin-infiltrating CD20(+) B cells display an atypical memory phenotype and correlate with favorable prognosis in hepatocellular carcinoma. *Clin cancer Res an Off J Am Assoc Cancer Res* (2013) **19**:5994–6005. doi:10.1158/1078-0432.CCR-12-3497
84. Fässler M, Diem S, Mangana J, Hasan Ali O, Berner F, Bomze D, Ring S, Niederer R, Del Carmen Gil Cruz C, Pérez Shibayama CI, et al. Antibodies as biomarker candidates for response and survival to checkpoint inhibitors in melanoma patients. *J Immunother Cancer* (2019) doi:10.1186/s40425-019-0523-2
85. Hamanaka Y, Suehiro Y, Fukui M, Shikichi K, Imai K, Hinoda Y. Circulating anti-MUC1 IgG antibodies as a favorable prognostic factor for pancreatic cancer. *Int J cancer* (2003) **103**:97–100. doi:10.1002/ijc.10801
86. Kurtenkov O, Klaamas K, Mensdorff-Pouilly S, Miljukhina L, Shljapnikova L, Chužmarov V. Humoral immune response to MUC1 and to the Thomsen-Friedenreich (TF) glycotope in patients with gastric cancer: Relation to survival. *Acta Oncol (Madr)* (2007) **46**:316–323. doi:10.1080/02841860601055441
87. Fremd C, Stefanovic S, Beckhove P, Pritsch M, Lim H, Wallwiener M, Heil J, Golatta M, Rom J, Sohn C, et al. Mucin 1-specific B cell immune responses and their impact on overall survival in breast cancer patients. *Oncoimmunology* (2015) **5**:e1057387–e1057387. doi:10.1080/2162402X.2015.1057387
88. Montfort A, Pearce O, Maniati E, Vincent BG, Bixby L, Böhm S, Dowe T, Wilkes EH, Chakravarty P, Thompson R, et al. A Strong B-cell Response Is Part of the Immune Landscape in Human High-Grade Serous Ovarian Metastases. *Clin cancer Res an Off J Am Assoc Cancer Res* (2017) **23**:250–262. doi:10.1158/1078-0432.CCR-16-0081
89. Clynes RA, Towers TL, Presta LG, Ravetch J V. Inhibitory Fc receptors modulate in vivo cytotoxicity against tumor targets. *Nat Med* (2000) **6**:443–446. doi:10.1038/74704
90. Cohen-Solal JFG, Cassard L, Fournier EM, Loncar SM, Fridman WH, Sautès-Fridman C. Metastatic melanomas express inhibitory low affinity fc gamma

- receptor and escape humoral immunity. *Dermatol Res Pract* (2010) **2010**:657406. doi:10.1155/2010/657406
91. Bolotin DA, Poslavsky S, Davydov AN, Frenkel FE, Fanchi L, Zolotareva OI, Hemmers S, Putintseva E V, Obraztsova AS, Shugay M, et al. Antigen receptor repertoire profiling from RNA-seq data. *Nat Biotechnol* (2017) **35**:908–911. doi:10.1038/nbt.3979
 92. Nicholas KJ, Zern EK, Barnett L, Smith RM, Lorey SL, Copeland CA, Sadagopal S, Kalams SA. B cell responses to HIV antigen are a potent correlate of viremia in HIV-1 infection and improve with PD-1 blockade. *PLoS One* (2013) **8**:e84185. doi:10.1371/journal.pone.0084185
 93. Ohaegbulam KC, Assal A, Lazar-Molnar E, Yao Y, Zang X. Human cancer immunotherapy with antibodies to the PD-1 and PD-L1 pathway. *Trends Mol Med* (2015) **21**:24–33. doi:10.1016/j.molmed.2014.10.009
 94. Diem S, Fässler M, Bomze D, Ali OH, Berner F, Niederer R, Hillmann D, Mangana J, Levesque MP, Dummer R, et al. Immunoglobulin G and Subclasses as Potential Biomarkers in Metastatic Melanoma Patients Starting Checkpoint Inhibitor Treatment. *J Immunother* (2019) **42**:89–93. doi:10.1097/CJI.0000000000000255
 95. Nguyen N, Mamchak A, Severgnini M, Williamson KS, Wu X, Drabek EF, Manos M, Chen X, Tang X, Amiri Z, et al. Abstract 615: Increased somatic hypermutation in the immunoglobulin sequences of melanoma patients who have durable response to checkpoint inhibitor therapy. in doi:10.1158/1538-7445.am2018-615
 96. Wu X, Li J, Connolly EM, Liao X, Ouyang J, Giobbie-Hurder A, Lawrence D, McDermott D, Murphy G, Zhou J, et al. Combined Anti-VEGF and Anti-CTLA-4 Therapy Elicits Humoral Immunity to Galectin-1 Which Is Associated with Favorable Clinical Outcomes. *Cancer Immunol Res* (2017) **5**:446–454. doi:10.1158/2326-6066.CIR-16-0385
 97. Buch MH, Boyle DL, Rosengren S, Saleem B, Reece RJ, Rhodes LA, Radjenovic A, English A, Tang H, Vratsanos G, et al. Mode of action of abatacept in rheumatoid arthritis patients having failed tumour necrosis factor blockade: A histological, gene expression and dynamic magnetic resonance imaging pilot study. *Ann Rheum Dis* (2009) **68**:1220–1227. doi:10.1136/ard.2008.091876
 98. Genovese MC, Becker J-C, Schiff M, Luggen M, Sherrer Y, Kremer J, Birbara C, Box J, Natarajan K, Nuamah I, et al. Abatacept for Rheumatoid Arthritis Refractory to Tumor Necrosis Factor α Inhibition. *N Engl J Med* (2005) **353**:1114–1123. doi:10.1056/nejmoa050524
 99. Kremer JM, Genant HK, Moreland LW, Russell AS, Emery P, Abud-Mendoza C, Szechinski J, Li T, Ge Z, Becker JC, et al. Effects of abatacept in patients with methotrexate-resistant active rheumatoid arthritis: A randomized trial. *Ann Intern Med* (2006) **144**:865–876. doi:10.7326/0003-4819-144-12-200606200-00003
 100. Linsley PS, Wallace PM, Johnson J, Gibson MG, Greene JAL, Ledbetter JA, Singh C, Tepper MA. Immunosuppression in vivo by a soluble form of the CTLA-4 T cell activation molecule. *Science* (80-) (1992) **257**:792–795. doi:10.1126/science.1496399
 101. Tay L, Leon F, Vratsanos G, Raymond R, Corbo M. Vaccination response to tetanus toxoid and 23-valent pneumococcal vaccines following administration of a single dose of abatacept: A randomized, open-label, parallel group study in healthy subjects. *Arthritis Res Ther* (2007) **9**: doi:10.1186/ar2174
 102. Osorio JC, Ni A, Chaft JE, Pollina R, Kasler MK, Stephens D, Rodriguez C, Cambridge L, Rizvi H, Wolchok JD, et al. Antibody-mediated thyroid dysfunction during T-cell checkpoint blockade in patients with non-small-cell lung cancer. *Ann Oncol* (2017) **28**:583–589. doi:10.1093/annonc/mdw640

103. Kobayashi T, Iwama S, Yasuda Y, Okada N, Tsunekawa T, Onoue T, Takagi H, Hagiwara D, Ito Y, Morishita Y, et al. Patients with antithyroid antibodies are prone to develop destructive thyroiditis by nivolumab: A prospective study. *J Endocr Soc* (2018) **2**:241–251. doi:10.1210/JS.2017-00432
104. Shirai T, Sano T, Kamijo F, Saito N, Miyake T, Kodaira M, Katoh N, Nishie K, Okuyama R, Uhara H. Acetylcholine receptor binding antibody-associated myasthenia gravis and rhabdomyolysis induced by nivolumab in a patient with melanoma. *Jpn J Clin Oncol* (2016) **46**:86–88. doi:10.1093/jjco/hyv158
105. Bluestone JA, Anderson M, Herold KC, Stamatouli AM, Quandt Z, Perdigo AL, Clark PL, Kluger H, Weiss SA, Gettinger S, et al. Collateral damage: Insulin-dependent diabetes induced with checkpoint inhibitors. in *Diabetes* (American Diabetes Association Inc.), 1471–1480. doi:10.2337/dbi18-0002
106. Kong BY, Micklethwaite KP, Swaminathan S, Kefford RF, Carlino MS. Autoimmune hemolytic anemia induced by anti-PD-1 therapy in metastatic melanoma. *Melanoma Res* (2016) **26**:202–204. doi:10.1097/CMR.0000000000000232
107. De Moel EC, Rozeman EA, Kapiteijn EH, Verdegaal EME, Grummels A, Bakker JA, Huizinga TWJ, Haanen JB, Toes REM, Van Der Woude D. Autoantibody development under treatment with immune-checkpoint inhibitors. *Cancer Immunol Res* (2019) **7**:6–11. doi:10.1158/2326-6066.CIR-18-0245
108. Cogdill AP, Andrews MC, Wargo JA. Hallmarks of response to immune checkpoint blockade. *Br J Cancer* (2017) **117**:1–7. doi:10.1038/bjc.2017.136
109. Jenkins RW, Barbie DA, Flaherty KT. Mechanisms of resistance to immune checkpoint inhibitors. *Br J Cancer* (2018) **118**:9–16. doi:10.1038/bjc.2017.434
110. Pinc A, Somasundaram R, Wagner C, Hörmann M, Karanikas G, Jalili A, Bauer W, Brunner P, Grabmeier-Pfistershammer K, Gschaider M, et al. Targeting CD20 in melanoma patients at high risk of disease recurrence. *Mol Ther* (2012) **20**:1056–1062. doi:10.1038/mt.2012.27
111. Josephs DH, Spicer JF, Karagiannis P, Gould HJ, Karagiannis SN. IgE immunotherapy: a novel concept with promise for the treatment of cancer. *MAbs* (2014) **6**:54–72. doi:10.4161/mabs.27029
112. Simpson TR, Li F, Montalvo-Ortiz W, Sepulveda MA, Bergerhoff K, Arce F, Roddie C, Henry JY, Yagita H, Wolchok JD, et al. Fc-dependent depletion of tumor-infiltrating regulatory T cells co-defines the efficacy of anti-CTLA-4 therapy against melanoma. *J Exp Med* (2013) **210**:1695–1710. doi:10.1084/jem.20130579
113. Quezada SA, Peggs KS. Lost in translation: Deciphering the mechanism of action of anti-human CTLA-4. *Clin Cancer Res* (2019) **25**:1130–1132. doi:10.1158/1078-0432.CCR-18-2509
114. Cabrita R, Lauss M, Sanna A, Donia M, Skaarup Larsen M, Mitra S, Johansson I, Phung B, Harbst K, Vallon-Christersson J, et al. Tertiary lymphoid structures improve immunotherapy and survival in melanoma. *Nature* (2020) **577**:561–565. doi:10.1038/s41586-019-1914-8

Table 1: Summary of current evidence of a humoral immune response in patients treated with checkpoint inhibitor therapy.

Sample type	Time point	CPI used	Target	B cell and antibody responses	Reference
Tumour	Baseline	Ipilimumab	CTLA-4	<ul style="list-style-type: none"> IL10+ Breg enrichment found in non-responders to anti-CTLA-4 therapy Absence of IGHG gene signature enriched in non-responders to anti-CTLA-4 	(22)
		Nivolumab	PD-1	<ul style="list-style-type: none"> IL10+ Breg presence did not correlate with treatment response to anti-PD-1 IGHG gene signature did not correlate with treatment response to anti-PD-1 	
Tumour	Baseline	Ipilimumab & Nivolumab combination	CTLA-4 & PD-1	<ul style="list-style-type: none"> For all treatments increased B cell frequency found in the tumours of responders to both combination therapy and anti-PD-1 monotherapy in both baseline and early on treatment samples For all treatments higher density of CD20+ B cells and TLS and a higher TLS:tumour area ratio in early on-treatment samples from responders versus non-responders For all treatments increased frequency of switched memory B cells found in responders at baseline and early on treatment 	(75)
		Nivolumab	PD-1		
	Early on treatment	Ipilimumab & Nivolumab combination	CTLA-4 & PD-1		
		Nivolumab	PD-1		
Tumour	Baseline	Ipilimumab	CTLA-4	<ul style="list-style-type: none"> B cells found to localise within TLS TLS gene signature present in responders" TLS structures associated with CD8+ T cells 	(114)
Peripheral blood	After 1 year of treatment	Ipilimumab	CTLA-4	<ul style="list-style-type: none"> High plasmablast numbers seen in patients responding to anti-CTLA-4 Sequenced plasmablast antibodies in patients who had not progressed after 1 year had somatic hypermutation, class switching and clonal expansion showing an active humoral response 	(78)
Tumour	Baseline	Nivolumab	PD-1	<ul style="list-style-type: none"> Plasmablast-like phenotype (CD19+CD20+CD38+CD138-) found to be most common B cell phenotype in TME Tumour induced plasmablast population (TIBP) defined using melanoma secretome Increased expression of TIBP gene signature in baseline samples associated with 	(41)

				improved response to Nivolumab	
Periphera l blood	Baseline	Nivolumab	PD-1	<ul style="list-style-type: none"> ▪ Melanoma-antigen specific antibodies to: MDA (TRP1/TYRP1, TRP2/TYRP2, gp100, MelanA/MART1) and the Cancer-Testis antigen NY-ESO-1 higher in responders to all treatments ▪ Melanoma-antigen specific IgGs higher in responders at baseline compared with non-responders for all treatments ▪ Melanoma-antigen specific IgGs in patients were mainly IgG1 and IgG2 	(84)
		Pembrolizumab	PD-1		
		Ipilimumab	CTLA-4		
		Ipilimumab & Nivolumab combination	CTLA-4 & PD-1		
Periphera l blood	Baseline	Nivolumab	PD-1	<ul style="list-style-type: none"> ▪ For all treatments high titres of IgG, IgG1, IgG2 and IgG3 showed a positive correlation with progression-free survival (PFS) ▪ For all treatments high titre of IgG2 showed a positive correlation with overall survival (OS) 	(Diem et al., 2019)
		Pembrolizumab	PD-1		
		Ipilimumab	CTLA-4		
		Ipilimumab & Nivolumab combination	CTLA-4 & PD-1		
Periphera l blood	Baseline	Ipilimumab & nivolumab combination therapy	CTLA-4 & PD-1	<ul style="list-style-type: none"> ▪ Overall decline in B cell numbers but increased plasmablast and CD21^{lo} B cell subsets corresponded to development of high grade immune related adverse events (irAE) ▪ Changes in other immune cells not significantly associated with frequency or severity of irAE 	(76)
	Early on-treatment				

Figure Legends

Figure 1 Proposed humoral responses in cancer and in response to checkpoint inhibitor immunotherapy.

Putative pro- and anti-tumour roles of B cells in the tumour microenvironment (TME) and tumour tertiary lymphoid structures (TLS). B cells may confer several anti-tumour roles through production of effective tumour-clearing antibodies, predominantly IgG1: mediating antibody dependent cell-mediated cytotoxicity (ADCC), antibody dependent cell-mediated phagocytosis (ADCP) and facilitating complement activation. B cells can also effectively modulate CD4 and CD8 T cells via their functions as antigen presenting cells (APCs) to present tumour-derived antigens or by promoting cytokine secretion, particularly IFN γ and IL-2 by T cells. B cells can also support the presentation of tumour-derived antigens by other antigen presenting cells including macrophages and dendritic cells. Antigen presentation and B cell activation may be enhanced in TLS, leading to somatic hypermutation, clonal amplification and isotype switching to activatory antibody isotypes such as IgG1, further driving anti-tumour responses. B cells can also directly exert cytotoxic functions against tumour cells (e.g. through secretion of granzyme B). B cells can have pro-tumour roles by multiple mechanisms, for instance through expression of inflammatory antibody isotypes such as IgG4. IL-10 producing regulatory B cell (Breg) phenotype is described which can negatively modulate the function of effector CD8 T cells, induce the development of regulatory T cells (Tregs) and dampen the activity of antigen-presenting cells such as dendritic cells and macrophages. IgA producing regulatory B cells have also been described in the TME.

Figure 2 Proposed B cell features at baseline that predict response to checkpoint inhibitor treatment.

B cell phenotypes enriched in tumours and the circulation of patients before treatment may be predictive of clinical response to checkpoint inhibitor therapy. Features include the presence of memory B cells (e.g. Ipilimumab monotherapy and combination of Ipilimumab and Nivolumab) and plasmablasts (e.g. Ipilimumab and Nivolumab monotherapies). In the peripheral blood, immunoglobulin isotypes IgG1, IgG2 and IgG3 have been associated with response to Ipilimumab and Nivolumab monotherapies. Melanoma-antigen specific IgG antibodies have been found to be increased in responders to therapy (monotherapy with Ipilimumab, Pembrolizumab and Nivolumab; as well as with combination of Ipilimumab and Nivolumab). Conversely, enrichment of tumours with IL10+ regulatory B cells and an absence of

an IGHG gene signature has been associated with a poor response to Ipilimumab.

FcγR: Fcγ receptor; NK: natural killer; TCR: T cell receptor.

Figure 1 Proposed roles of B cell in the tumour microenvironment

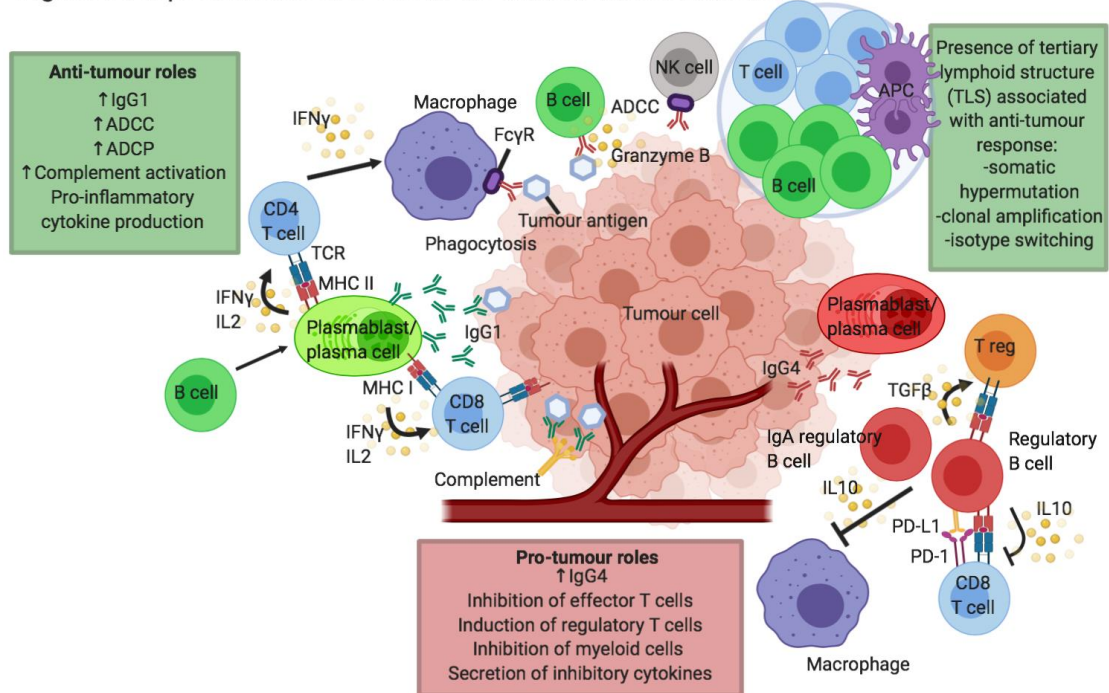


Figure 2 Proposed B cell changes at baseline that predict response to treatment

